



## ANTIMICROBIAL PACKAGING WITH NATURAL COMPOUNDS – A REVIEW

Renata Dobrucka

Poznan University of Economics, Poznan, **Poland**

**ABSTRACT. Background:** Packaging problems are an integral part of logistics and the implementation of packaging significantly affects the effectiveness of logistics processes, as a factor which increases the safety and the quality of products being transported. Active packaging is an area of technology needed to meet the requirements of the contemporary consumer. Active packaging creates additional opportunities in systems for packing goods, as well as offering a solution in which the packaging, the product and surroundings interact. Furthermore, active packaging allows packaging to interact with food and the environment and play a dynamic role in food preservation. The main role of antimicrobial packaging is to inhibit the growth of microorganisms that reduce the quality of the packaged product.

**Methods:** The application of natural antimicrobial agents appears to be safe for food products. Also, these compounds have potential applications as a natural preservative in the food packaging industry. This study presents some antibacterial agents, namely chitosan, nisin and pectins.

**Results and conclusion:** Natural substances used in active packaging can eliminate the danger of chemical substances migrating to food.

**Key words:** logistics, antimicrobial packaging, natural antibacterial agents.

### INTRODUCTION

The principal function of packaging is protection from heat, light, the presence or absence of moisture, oxygen, pressure, enzymes, microorganisms, insects, dirt and dust particles, gaseous emissions, and other things which might damage the product [Brody et al. 2001]. Antimicrobial packaging is a promising form of active packaging. Active packaging refers to the incorporation of certain additives into packaging film or within packaging containers, with the aim of maintaining and extending product shelf life [Day 1989]. Moreover, active packaging, sometimes referred to as interactive packaging, is intended to sense internal or external environmental change and to respond by

changing its own properties and hence the internal package environment [Brody et al., 2001].

Antimicrobial agents may be incorporated directly into packaging materials for slow release into the food surface or may be used in vapour form [Wilson 2007]. Antimicrobials reduce the growth rate and maximum population of microorganisms (spoilage and pathogenic) by extending the lag phase of microbes or inactivating them [Quintavalla, Vicini 2002]. The use of packaging films containing antimicrobial agents could be more efficient than the direct addition of these compounds into food. The nature of the antimicrobial agents is very diverse [organic acids, enzymes, bacteriocins, fungicides, natural extracts, ions, ethanol etc.] as well as

the nature of the materials into which they are included, such as papers, plastics, metals or combinations of these materials [Dainelli et al., 2008].

## ANTIMICROBIAL AGENTS

According to Nilsson et al. [2000] antimicrobial packaging technology that has the potential to modify headspace atmosphere creates an effective hurdle against bacterial cells. An antimicrobial in gaseous form is evenly distributed in the headspace of a packaging structure and functions more effectively compared to antimicrobials in solid or liquid forms. The most commonly used antimicrobial gas in food application is CO<sub>2</sub>, which inhibits both Gram-positive and Gram-negative microorganisms. Also, an alternative method of generating antimicrobial gas would be the removal of molecular O<sub>2</sub> from the headspace of a packaging structure. Molecular O<sub>2</sub> encourages the growth of obligate aerobic, facultative anaerobic, and microaerophilic microorganisms and causes the development of off flavor and off-color, as well as the loss of nutritional components of packaged food products [de Kruijf et al. 2002].

Antimicrobial effects which are achieved by adding antimicrobial agents into the packaging system or using antimicrobial polymeric materials generally function in one of three ways: release, absorption or immobilisation [Han 2003]. Also, the choice of an antimicrobial agent depends primarily on its activity against the targeted microorganisms [Han, 2000 2005]. Many other factors, however, need to be considered when designing antimicrobial packaging systems, such as specific activity, resistance of microorganisms, controlled release mechanisms, the chemical nature of foods and antimicrobials, storage and distribution conditions, film/container casting process conditions, physical and mechanical properties of antimicrobial packaging materials, organoleptic characteristics and toxicity of antimicrobials, and corresponding regulations [Han 2000]. All these factors should be carefully considered according to the corresponding regulations in order to design an effective active packaging [Sadaka et al. 2014].

As a result, there are many antimicrobial agents that exist and are widely used. Antimicrobial agents have different activities and affect different microorganisms. This is due to the characteristic antimicrobial mechanisms and also to the various physiologies of microorganisms. Therefore three groups of antimicrobial agents can be used: chemical agents, natural agents, and probiotics. Antibacterial agents of natural origin include chitosan, nisin and pectins.

## CHITOSAN

Chitosan is natural antimicrobial polymer obtained by deacetylation of chitin obtained commercially from shrimp and crab shell. Also, in literature chitosan has been reported as an antibacterial agent against a wide variety of microorganisms [Entsar et al. 2003, Wu et al. 2005]. The antimicrobial mechanism for chitosan is related to interactions of the cationic chitosan with the anionic cell membranes, increasing membrane permeability and eventually resulting in rupture and leakage of the intracellular material. Moreover, it has been reported that bulk chitosan and its nanoparticles are ineffective at pH<6, probably because of the absence of protonated amino groups [Qi et al. 2004]. The applications of chitosan in food packaging are mainly justified by their antimicrobial and antifungal activities against pathogenic and spoilage microbes. Chitosan-based active films allow food preservation to be extended and the use of chemical preservatives to be reduced [Aider 2010]. According to Rabea et al. [2003], chitosan exhibits greater antimicrobial activity than chitin, due to the greater number of free amino groups, which respond to the antimicrobial activity upon protonation. Cellulose and chitosan are the two most abundant biorenewable natural materials and have shown great promise for their application in food packaging. These naturally-occurring polymers have the ability to form films and moderate oxygen and moisture permeability [Byun et.al. 2012, Mi et al. 2006, Pereda et al 2011, Souza et al. 2010, Vargas et al. 2011]. Yu et al. [2013] prepared packaging films from water-soluble chitosan [N,O-carboxymethyl chitosan, NOCC] and cellulose [methylcellulose, MC]. They prepared active

packaging films with caffeic acid which was incorporated into the composite films in fixed and releasable types. The caffeic acid-incorporated films showed 20-fold increases in antioxidant activity and 6-fold increases in antibacterial activity as compared to the caffeic acid-free composite films. The releasable caffeic acid could be continuously released from the composite films and showed a significant inhibitory effect on lipid oxidation of menhaden oil-in-water emulsion. The chitosan antifungal action was evaluated against *Alternaria alternata*, *Aspergillus niger*, and *Rhizopus oryzae* [Ziani et al. 2009]. Cruz-Romero et al. [2013] presented antimicrobial activity of chitosan, organic acids and nano-sized solubilisates for potential use in smart antimicrobially-active packaging for potential food applications. Abdou et al. [in press] demonstrated that a plasma pretreated polypropylene surface treated with chitosan could impart antibacterial and antifungal properties, since chitosan has been shown to possess efficient antimicrobial abilities.

One of the most promising active bio-films is the one based on chitosan combined with different materials, such as plant and animal proteins, polysaccharides and antimicrobial peptides [bacteriocin] such as nisin and divergicin, a new class of bacteriocin produced by *Carnobacterium divergens* [Tahiri et al. 2004, Tahiri et al. 2009]. Moreover, Elsabee et al. [2008] studied tomato packing in bags of transparent polypropylene film coated with twelve non-nanoscale alternating layers of chitosan and pectin, in view of a new concept of active packaging for fruit preservation. Chitosan and pectin can interact at pH 5.6. However the gel behaviour depends upon the degree of esterification of the pectin. In fact the polyelectrolyte complex formation requires ionized carboxylate groups of pectin and protonated amino groups of chitosan [Lehr et al. 1992]. Wang et al. [2015] prepared antibacterial packaging film containing chitosan/poly[vinyl alcohol] [Nisin-CS/PVA]. The experimental results showed antimicrobial activity films against *S. aureus* which may have potential as an active film in food packaging. Duran et al. [2016] presented the use of nisin, natamycin, pomegranate and grape seed extracts in chitosan coating to extend the shelf life of strawberries.

Antimicrobial agents were added to chitosan at a concentration of 1% w/v. The results showed that all coatings have a good effect on the quality of strawberries during the storage period.

## NISIN

Nisin is an antimicrobial peptide with 34 amino acids and a molecular weight of 3.5 kDa, produced by strains of *Lactococcus lactis* subsp. *lactis* [Mulders et al. 1991]. Moreover, is the most common bacteriocin, tested for many applications. This peptide has antimicrobial properties, especially against the food-borne pathogens *Listeria monocytogenes*, *Staphylococcus aureus* or *Bacillus cereus* [Brewer et al. 2002, Lopez-Pedemonte et al. 2003]. Due to its effect on the important Gram positive food-borne pathogens, many studies have focused on the incorporation of nisin into various kinds of films made of cellulose, nylon, whey protein isolate, hydroxypropyl methylcellulose, zein etc. and their use as nisin delivery system packaging films to reduce undesirable bacteria in foodstuffs [Chollet et al. 2008, Coma et al. 2001, Gadang et al. 2008, Ko et al. 2001, Kristo et al. 2008, Natrajan Sheldon 2000, Neetoo et al. 2008, Nguyen et al. 2008, Teerakarn et al. 2002]. Studies have also been published by a number of authors on the use of nisin as an antimicrobial in a wide variety of food products [Delves-Broughton et al. 1996]. Nguyen et al. [2008] prepared nisin-containing bacterial cellulose film to inhibit *Listeria monocytogenes* on processed meats. This cellulose film containing nisin was developed and used in a proof-of-concept study to control *Listeria monocytogenes* and total aerobic bacteria on the surface of vacuum-packed frankfurters. Additionally, many studies have shown that nisin may be efficiently incorporated into cellulose-based packaging films and used for controlling pathogens in food products [Ming et al. 1997, Scannell et al. 2000, Franklin et al. 2004, Luchansky & Call 2004].

Ercolini et al. [2010] studied the spoilage-related microbial populations of beef and to investigate the effect of nisin-activated antimicrobial packaging on the development of beef spoilage at low temperatures.

The combination of chill temperatures and antimicrobial packaging proved to be effective in enhancing the microbiological quality of beef cuts by inhibiting LAB, carnobacteria and *Brochothrix thermosphacta* in the early stages of storage and by reducing the loads of Enterobacteriaceae. Economou et al. [2009] presented the effect of nisin and EDTA treatments in increasing the shelf-life of raw poultry products stored under modified atmosphere packaging at 4°C. Neetoo et al. [2008] used nisin-coated plastic films to control *Listeria monocytogenes* on vacuum-packaged cold-smoked salmon. The effect of storage temperature, nisin concentration on low-density polyethylene [LDPE] film, and inoculation level on the growth and survival of *Listeria monocytogenes* was investigated. At 4°C [low and high inoculum levels] and 10 °C [low inoculum level], it was found that the degree of inactivation or growth inhibition of *Listeria monocytogenes* was directly related to the concentration of nisin. The fact that nisin delayed the growth of *Listeria monocytogenes* populations in smoked salmon at both low and high inoculum levels show that nisin might be used to control post-processing contamination of *Listeria monocytogenes* in cold-smoked salmon. Cao-Hoang et al. [2010] presented the effectiveness of the nisin-coated sodium caseinate films against *Listeria innocua* in cheese during storage at refrigerated temperatures, indicating that combining nisin into sodium caseinate films is a promising method to enhance the safety and extend the shelf life of processed cheeses.

## PECTIN

Pectin is a natural polysaccharide, poly [1,4-galacturonic acid], which is obtained from the cell walls of terrestrial plants and exhibits polyanionic behaviour [Farris et al. 2011, May 1990]. It is now known that pectin is a major component of the plant cell wall and the most complex macromolecule in nature [Voragen et al., 2009]. Pectin is also a high-molecular weight, biocompatible, non-toxic, and anionic natural polysaccharide extracted from cell walls of higher plants [Zouambia et al. 2009]. Gopi et al. [2014] extracted pectin from banana peel. The experimental results revealed that the hydroxyapatite HAP nanoparticles synthesized

in the presence of an optimized concentration of pectin are pure, low crystalline, spherical and discrete particles with reduced size. Ravishankar et al. [2012] studied films based on pectin and apple, carrot or hibiscus. The films were treated with carvacrol or cinnamaldehyde and their antimicrobial activity was tested against *Listeria monocytogenes* on contaminated ham and bologna.

Physical or chemical modification of pectin can lead to new products with significant functional properties. The applications of pectin have been also extended greatly from food and food additives to various fields, such as drug delivery [Mishra et al. 2008, Souto-Maior et al. 2010], antithrombotic agents [Cipriani et al. 2009], and mucoadhesive [Sharma, Ahuja 2011] and antimicrobial substances in materials for packaging food. Moreover, Tripathi et al. [2010] developed an antimicrobial chitosan/poly [vinyl alcohol]/pectin ternary film for food packaging applications. It exhibited significant antimicrobial activity against various pathogenic bacteria such as *Escherichia coli*, *Staphylococcus aureus*, *Bacillus subtilis*, *Pseudomonas* and *Candida albicans*. Pectin is a general term for a group of valuable natural polysaccharides extracted from edible plant material where they occur as structural materials. Its main sources are citrus peel and apple pomace [Stasse-Wolthuis et al. 1980, Thibault, Ralet 2001]. Gorrasi et al. [2012] prepared films containing pectins obtained from apples. The films showed antimicrobial activity, indicating the potential application of prepared complexes in the packaging field and the potential usage of pectin-antimicrobials as coating agents for a wide number of packaging polymers. The antimicrobial activity of pectin edible films incorporated with nisin and its combination with treatment of ionizing radiation was used to control *Listeria monocytogenes* on a ready-to-eat [RTE] turkey meat by Jin et al. [2009]. The combination of irradiation with pectin film containing nisin resulted in a 3.95 log CFU/cm<sup>2</sup> reduction at 1 kGy and a 5.35 log CFU/cm<sup>2</sup> reduction at 2 kGy; indicating a synergistic effect on *Listeria* viability on the surface of RTE turkey meat. In the same year, Jin et al. [2009] tested pectin-poly lactide [PLA] composite films

treated with with nisin against *Listeria monocytogenes*. Alves et al. [2011] developed model composite films based on commercial pectin and carrageenan, containing organically modified nanoclays. The barrier properties to water vapour and CO<sub>2</sub> of a polymeric matrix composed by kappa-carrageenan and pectin [66.7% kappa-carrageenan], with the inclusion of nanoclays, was studied. The effect of particle content of the films on water vapour permeability [WVP] was dependent on the driving force applied.

## SUMMARY

Traditional food packaging is designed to mechanically support otherwise non-solid food, and to protect food from external influences. Antimicrobial packaging appears to be one of the most promising applications of active food packaging technology.

Natural compounds that do not have any significant medical or environmental impact could potentially serve as effective alternatives to conventional antibacterial or antifungal agents. Therefore, the preparation of antimicrobial materials with natural substances is a huge challenge to researchers and producers of packaging food.

## REFERENCES

- Aider M., 2010. Chitosan application for active bio-based films production and potential in the food industry: Review, *Food Science and Technology*, 43, 837-842.
- Ali S.W., Rajendran S., Joshi M., 2011. Synthesis and characterization of chitosan and silver loaded chitosan nanoparticles for bioactive polyester. *Carbohydrate Polymers*, 83, 438-446.
- Alves V.D., Castelló R., Ferreira A.R., Nuno Costa N., Fonseca I.M., Coelho I. M., 2011. Barrier properties of carrageenan/pectin biodegradable composite films. *Procedia Food Science*, 1, 240 - 245.
- Brewer R., Adams M.R., Park S.F. 2002. Enhanced inactivation of *Listeria monocytogenes* by nisin in the presence of ethanol. *Lett. Appl. Microbiol.*, 34, 18-21.
- Brody A, Strupinsky ER, Kline LR. 2001. Odor removers. In: Brody A, Strupinsky ER, Kline LR, editors. *Active packaging for food applications*. Lancaster, Pa.: Technomic Publishing Company, Inc. 107-17.
- Byun Y., Ward A., Whiteside S., 2012. Formation and characterization of shellac-hydroxypropyl methylcellulose composite films. *Food Hydrocolloids*, 27, 364-370.
- Cao-Hoang L., Chaine A., Grégoire L, Waché Y. 2010. Potential of nisin-incorporated sodium caseinate films to control *Listeria* in artificially contaminated cheese. *Food Microbiology*, 27, 940-944.
- Chen H.B., Chiou B.S., Wang Y.Z., Schiraldi D. A. 2013. Biodegradable pectin/clay aerogels. *ACS Applied Materials & Interfaces*, 5[5], 1715-1721.
- Chollet E., Sebti I., Martial-Gros A., Degraeve P., 2008. Nisin preliminary study as a potential preservative for sliced ripened cheese: NaCl, fat and enzymes influence on nisin concentration and its antimicrobial activity. *Food Control*, 19, 982-989.
- Cipriani T.R., Gracher A.H.R., de Souza L.M., Fonseca R.J.C., Belmiro C.L.R., Gorin P.A.J., 2009. Influence of molecular weight of chemically sulfated citrus pectin fractions on their antithrombotic and bleeding effects. *Thrombosis and Haemostasis*, 101[5], 860-866.
- Coma V., Sebti I., Pardon P., Deschamps A., Pichavant F.H. 2001. Antimicrobial edible packaging based on cellulosic ethers, fatty acids and nisin incorporation to inhibit *Listeria innocua* and *Staphylococcus aureus*. *J Food Prot*, 64, 470-475.
- Coma V., 2008. Bioactive packaging technologies for extended shelf life of meatbased products. *Meat Science*, 78, 90-103.
- Cruz-Romero M.C., Murphy T., Morris M., Cummins E., Kerry J.P. 2013. Antimicrobial activity of chitosan, organic acids and nano-sized solubilisates for potential use in smart antimicrobially-active packaging for potential food applications, *Food Control*, 34, 393-397.

- Dainelli D., Gontard N., Spyropoulos D., Zondervan-van den Beuken E., Tobback P. 2008. Active and intelligent food packaging: legal aspects and safety concerns. *Trends in Food Science & Technology*, 19, 99 - 108.
- Day, B.P.F., 1989, Extension of shelf-life of chilled foods. *Eur Food Drink Rev.* 4, 47-56.
- Delves-Broughton J., Blackburn P., Evans R. J., Hugenholz J. 1996. Applications of the bacteriocin nisin. *Antonie van Leeuwenhoek*, 69, 193-202.
- Duran M., Seckin Aday M., Zorba N.N.D., Temizkan R., Büyükcan M.B., Caner C., 2016. Potential of antimicrobial active packaging'containing natamycin, nisin, pomegranate and grape seed extract in chitosan coating' to extend shelf life of fresh strawberry. *Food and Bioprocess Technology*, 98 (2016) 354-363.
- Economou T., Pournis N., Ntzimani A., Savvaidis I.N. 2009. Nisin-EDTA treatments and modified atmosphere packaging to increase fresh chicken meat shelf-life. *Food Chemistry*, 114, 1470-1476.
- Elsabee M.Z., Abdou E.S., Nagy K.S.A. 2008. Eweis M. Surface modification of polypropylene films by chitosan and chitosan/pectin multilayer. *Carbohydrate Polymers*, 71[2], 187-195.
- Entsar I.R., Badawy M.E.T., Stevens C.V., Smaghe G., Walter S., 2003. Chitosan as antimicrobial agent: application and mode of action. *Biomacromolecules*, 4, 1457-1465.
- Ercolini D., Ferrocino I., La Stora A., Mauriello G., Gigli S., Masi P., Francesco Villani F. 2010. Development of spoilage microbiota in beef stored in nisin activated packaging *Food Microbiology*, 27,137-143.
- Farris S., Schaich K.M., Liu L.S., Cooke P.H., Piergiovanni L., Yam, K.L., 2011. Gelatine pectin composite films from polyion-complex hydrogels. *Food Hydrocolloid*, 25, 61 -70.
- Floros J.D., Dock L.L., Han J.H., 1997. Active packaging technologies and applications. *Food Cosmetics and Drug Packaging*, [S.I.], 17, 10.
- Franklin N.B., Cooksey K.D., Getty K.J.K., 2004. Inhibition of *Listeria monocytogenes* on the surface of individually packaged hot dogs with a packaging film coating containing nisin. *J. Food Prot.* 67, 480-485.
- Gadang V.P., Hettiarachchy N.S., Johnson M.G., Owens C., 2008. Evaluation of antibacterial activity of whey protein isolate coating incorporated with nisin, grape seed extract, malic acid, and EDTA on a Turkey frankfurter system. *J. Food Sci.*, 73, 389-394.
- Gopi D., Shinyjoy E., Kavitha L., 2014. Synthesis and spectral characterization of silver/magnesium co-substituted hydroxylapatite for biomedical applications, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 118, 589-597.
- Gontard N., 2007. Antimicrobial paper based packaging. In: *International antimicrobial in plastic and textile applications. InterTech PIRA conference. 27-26 June 2007, Prague, Czech Republic.*
- Gorrasi G., Bugatti V., Vittoria V., 2012. Pectins filled with LDH-antimicrobial molecules: Preparation, characterization and physical properties. *Carbohydrate Polymers*, 89, 132- 137.
- Han J.H., 2000, *Antimicrobial Food Packaging*. *Food Technology*, 54[3], 56-65.
- Han J.H., 2003. *Antimicrobial food packaging* In R. Ahvenainen [Ed.] *Novel food packaging techniques: Woodhead Publishing Limited and CRC Press LLC*, 70-72.
- Han J.H., 2005. *Antimicrobial packaging systems*. In J.H. Han [Ed.], *Innovations in food packaging: Academic Press Inc.* 80-201.
- Ko S., Janes M.E., Hettiarachchy N.S., Johnson, M.G., 2001. Physical and chemical of edible films containing nisin and their action against *Listeria monocytogenes*. *Journal of Food Science*, 66[7], 1006-1011.

- Kristo E., Koutsoumanis K.P., Biliaderis C.G., 2008. Thermal, mechanical and water vapor barrier properties of sodium caseinate films containing antimicrobials and their inhibitory action on *Listeria monocytogenes*. *Food Hydrocolloids*, 22[3], 373-386.
- de Kruijf N., van Beest M., Rijk R., Sipilainenmalm T., Paseiro Losada P., De Meulenaer B. 2002. Active and intelligent packaging: applications and regulatory aspects. *Food Additives & Contaminants*, 19, 144-162.
- Jin T., Liu L., Sommers C.H., Boyd G., Zhang, H., 2009. Radiation sensitization and postirradiation proliferation of *Listeria monocytogenes* on ready-to-eat deli meat in the presence of pectin nisin films. *Journal of Food Protection*. 72[3], 644-649.
- Qi L., Xu, Z., Jiang X., Hu C., Zou, X., 2004. Preparation and antibacterial activity of chitosan nanoparticles. *Carbohydrate Research*, 339[16], 2693-2700.
- Quintavall S., Vicini L., 2002, Antimicrobial food packaging in meat industry. *Meat Science*, 62, 373-380.
- Lehr C.M., Bouwstra J.A., Schacht E.H., Jungiger, H.E., 1992. In vitro evaluation of mucoadhesive properties of chitosan and some other natural polymers. *International Journal of Pharmaceutics*, 78[1-3],43-48.
- Lopez-Pedemonte T.J., Roig-Sagues A.X., Trujillo A.J., Capellas M., Guamis. B., 2003. Inactivation of spores of *Bacillus cereus* in cheese by high hydrostatic pressure with the addition of nisin or lysozyme. *J. Dairy Sci.*, 86, 3075-3081.
- Luchansky J.B., Call J.E., Hristova B., Rumery L., Yoder, L., Oser A. 2005. Viability of *Listeria monocytogenes* on commercially-prepared hams surface treated with acidic calcium sulfate and lauric arginate and stored at 4°C. *Meat Science*, 71, 92-99.
- May C.D., 1990. Industrial pectins: Sources, production, and applications. *Carbohydrate Polymers*, 12[1], 79-99.
- Mi F.L., Yu S.H., Peng C.K., Sung H.W., Shyu S.S., Liang H.F. Huang M.F., Wang C., 2006. Synthesis and characterization of a novel glycoconjugated macromolecule. *Polymer*, 47, 4348-4358.
- Ming X., Weber G.H., Ayres J.W., Sandine W.E., 1997. Bacteriocins applied to food packaging materials to inhibit *Listeria monocytogenes* on meats. *J. Food Sci.* 62, 413-415.
- Mishra R.K., Banthia A.K., Majeed A.B.A. 2012. Pectin based formulations for applications: a review. *Asian Journal of Pharmaceutical and Clinical Research*, 5[4], 1-7.
- Mulders J.W., Boerrigter I.J., Rollema H.S., Siezen R.J., de Vos W.M., 1991, Identification and characterization of the lantibiotic nisin Z, a natural nisin variant. *Eur J. Biochem.*, Nov 1, 201, 3, 581-584.
- Natrajan N., Sheldon B.W. 2000. Inhibition of *Salmonella* on poultry skin using protein- and polysaccharide-based films containing a nisin formulation. *J. Food Prot.*, 63, 1268-1272.
- Neetoo, H., Ye, M., Chen, H., Joerger, R.D., Hicks, D., Hoover, D.G., 2008. Use of nisin coated plastic films to control *Listeria monocytogenes* on vacuum packaged cold smoked salmon. *Int. J. Food Microbiol.*, 22, 8-15.
- Nguyen V.,T., Gidley M.J., Dykes, G., 2008. A.Potential of nisin-containing bacterial cellulose film to inhibit *Listeria monocytogenes* on processed meats. *Food Microbiology*, 25 [3], 471-478.
- Nilsson L., Chen Y., Chikindas M.L., Huss H., H., Gram L., Montville T.J., 2000. Carbon dioxide and nisin act synergistically on *Listeria monocytogenes*. *Applied Environmental Microbiology*, 66, 769-774.
- Park S.Y., Jun S.T., Marsh K.S., 2001. Physical properties of PVOH/chitosan blended films cast from different solvents. *Food Hydrocolloids*, 15[4-6], 499-502.
- Pereda M., Ponce A.G., Marcovich N.E., Ruseckaite R.A. Martucci J.F.C., 2011. Hitosan-gelatin composites and bi-layer films with potential antimicrobial activity. *Food Hydrocolloids*, 25, 1372-1381.
- Ravishankar S., Jaroni D., Zhu L., Olsen C., McHugh T., Friedman M., 2012. Inactivation of *Listeria monocytogenes* on ham and bologna using pectin-based apple, carrot, and hibiscus edible films containing

- carvacrol and cinnamaldehyde. *Journal of Food Science*, 77[7], 377-382.
- Rabea E.I., Badawy M.E., Stevens C.V., Smaghe G., Steurbaut W. 2013. Chitosan as antimicrobial agent: applications and mode of action. *Biomacromolecules*, 4, 1457-1465.
- Rhim J.W., Hong S.I., Park H.M., Ng P.K.W., 2006: Preparation and characterization of chitosan-based nanocomposite films with antimicrobial activity. *Journal of Agricultural and Food Chemistry*, 54, 5814-5822.
- Robertson, G.L. [ed.], 2006. *Food Packaging - Principles and Practice*. Second edition, CRC Press, Boca Raton, FL, USA.
- Scannell A.G.M., Hill C., Ross R.P., Marx S., Hartmeier W., Arendt E.K., 2000. Development of bioactive food packaging materials using immobilised bacteriocins Lacticin 3147 and Nisaplin [R]. *Int. J. Food Microbiol.* 2000, 60, 241-249.
- Souto-Maior J.F.A., Reis A.V., Pedreiro L.N., Cavalcanti O.A., 2010. Phosphated cross-linked pectin as a potential excipient for specific drug delivery: Preparation and physicochemical characterization. *Polymer International*, 59[1], 127-135.
- Souza M.P., Cerqueira M.A., Souza B.W.S., Teixeira J.A., Porto A.L.F., Vicente A.A. 2010. Polysaccharide from *Anacardium occidentale* L. tree gum [Policaju] as a coating for Tommy Atkins mangoes. *Chemical Papers*, 64[4], 475-481.
- Sharma R., Ahuja, M., 2011. Thiolated pectin: Synthesis, characterization and evaluation as a mucoadhesive polymer. *Carbohydrate Polymers*, 2011, 85[3], 658-663.
- Siragusa G.R., Cutter C.N., Willett J.L., 1999. Incorporation of bacteriocin in plastic retains activity and inhibits surface growth of bacteria on meat. *Food Microbiol.* 16, 229-235.
- Stasse-Wolthuis M., Albers H.F., van Jeveren J.G., Wil de Jong J., Hautvast J.G., 1980. Hermus R. J. Influence of dietary fiber from vegetables and fruits, bran or citrus pectin on serum lipids, fecal lipids, and colonic function. *American Journal of Clinical Nutrition*, 33, 1745-1756.
- Tahiri I., Desbiens M., Benech R., Kheadr E., Lacroix C., Thibault S., 2004. Purification, characterization and amino acid sequencing of divergicin M35: a novel class IIa bacteriocin produced by *Carnobacterium divergens* M35. *International Journal of Food Microbiology*, 97[2], 123-136.
- Tahiri I., Desbiens M., Lacroix C., Kheadr E., Fliss I., 2009. Growth of *Carnobacterium divergens* M35 and production of Divergicin M35 in snow crab by-product, a natural-grade medium. *LWT - Food Science and Technology*, 42[2], 624-632.
- Teerakarn A., Hirt D.E., Acton J.C., Rieck J.R., Dawson P.L. 2002. Nisin Diffusion in rotein Films: Effects of Film Type and Temperature, *Journal of Food Science*. 67, 8, 3019-3025.
- Thibault J.F., Ralet, M.C. 2001. Pectins, their origin, structure and functions. In B. V. McCleary, L. Prosky [Eds.], *2Advanced dietary fibre technology* [pp. 369-378]. Oxford, UK: Blackwell Science.
- Tripathi S., Mehrotra G.K., Dutta P.K., Preparation and physicochemical evaluation of chitosan/poly(vinyl alcohol)/pectin ternary film for food-packaging applications. *Carbohydrate Polymers*, 79[3], 711-716.
- Wang Y., Zhang Q., Zhang C., Li P., 2012. Characterisation and cooperative antimicrobial properties of chitosan/nano-ZnO composite nanofibrous membranes. *Food Chemistry*, 2012, 132, 419-427.
- Wang H., Zhang R., Zhang H., Jiang S., Liu H., Sun M., Jiang S. 2015. Kinetics and functional effectiveness of nisin loaded antimicrobial packaging film based on chitosan/poly(vinyl alcohol). *Carbohydrate Polymers* 127 [2015] 64-71.
- Wilson C., 2007. *Frontiers of intelligent and active packaging for fruits and vegetables*. Boca Raton, Fa.: CRC Press. 360.
- Wu T., Zivanovic S., Draughon F.A., Conway W.S., Sams C.E., 2005. Physicochemical properties and bioactivity of fungal chitin and chitosan. *Journal of Agricultural and Food Chemistry*, 53, 3888-3894.
- Vargas M., Albors A., Chiralt A., Gonzalez-Martinez C., 2006. Quality of cold-stored strawberries as affected by chitosan-oleic

- acid edible coatings. *Postharvest Biology and Technology*, 41, 164-171.
- Voragen A.G.J., Coenen G.J., Verhoef R.P., Schols H.A., 2009. Pectin, a versatile polysaccharide present in plant cell walls. *Structural Chemistry*, 20, 263-275.
- Yu S.H., Hsieh H.Y., Pang J.Ch., Tang D.W., Shih C.M., Tsai M.L., Tsai Y.C., Mi F.L., 2013. Active films from water-soluble chitosan/cellulose composites incorporating releasable caffeic acid for inhibition of lipid oxidation in fish oil emulsions. *Food Hydrocolloids*, 32, 9-19.
- Ziani K., Fernandez-Pan I., Royo M. Mate J.I., 2009. Antifungal activity of films and solutions based on chitosan against typical seed fungi. *Food Hydrocolloids*, 23, 2309-2314.
- Zouambia Y., Moulai- Mostefa N., Krea M., 2009. Structural characterization and surface activity of hydrophobically functionalized extracted pectins. *Carbohydrate Polymers*, 78, 841-846.
- Zhou Y., Zhao Y., Wang L., Xu L., Zhai M., Wei S. 2012. Radiation synthesis and characterization of nanosilver/gelatin/carboxymethyl chitosan hydrogel. *Radiation Physics and Chemistry*, 81, 553-56.

## AKTYWNE OPAKOWANIA Z NATURALNYMI ZWIĄZKAMI ANTYBAKTERYJNYMI

**STRESZCZENIE. Wstęp:** Problematyka opakowań jest elementem logistyki, zaś zastosowane opakowania istotnie wpływają na efektywność procesów logistycznych, jako czynnik zapewniający bezpieczeństwo i zwiększający jakość transportowanych wyrobów. Opakowania aktywne są obszarem technologii wychodzącym na przeciw wymaganiom stawianym przez współczesnego konsumenta. Opakowania aktywne stwarzają nowe możliwości w zakresie systemów pakowania towarów oraz stanowią rozwiązanie, w którym to opakowanie, produkt oraz otoczenie wzajemnie na siebie oddziałują. Poza tym opakowania aktywne w wyniku zachodzących oddziaływań z wewnętrzną atmosferą i produktem prowadzą do przedłużenia jego trwałości. Główną rolą opakowania przeciwbakteryjnego jest hamowanie wzrostu drobnoustrojów, które obniżają jakość produktu.

**Metody:** Stosowanie naturalnych środków przeciwbakteryjnych wydaje się być bezpieczne dla produktów spożywczych. Ten typ związków ma potencjalne zastosowanie jako naturalne środki konserwujące w przemyśle spożywczym. W niniejszej pracy przedstawiono niektóre ze stosowanych środków przeciwbakteryjnych, min. chitozan, nizyny i pektyny.

**Wyniki i podsumowanie:** Naturalne substancje przeciwbakteryjne stosowane w opakowaniach aktywnych eliminują niebezpieczeństwo konsumentów w zakresie migracji chemicznych substancji do żywności.

**Słowa kluczowe:** logistyka, opakowania antybakteryjne, naturalne substancje antybakteryjne

## ANTIBAKTERIELLE VERPACKUNGEN MIT NATÜRLICHEN VERBINDUNGEN

**ZUSAMMENFASSUNG. Einleitung:** Aktive Verpackungen stellen einen Technologie-Bereich dar, der den Anforderungen der gegenwärtigen Verbraucher gerecht zu werden vermag. Die aktiven Verpackungssysteme schaffen neue Möglichkeiten auf dem Gebiet des Verpackens von Waren und somit bilden sie eine Lösung, innerhalb deren die Verpackung, das Produkt und die Umwelt in Wechselwirkung miteinander treten. Darüber hinaus führen die aktiven Verpackungen infolge der aktiven Wechselwirkung der Innenatmosphäre mit dem betreffenden Produkt zur Verlängerung dessen Lebensdauer. Die Hauptrolle der Anwendung von antibakteriellen Verpackungen beruht auf der Hemmung des Wachstums von Mikroorganismen, die die Qualität des Produktes vermindern.

**Methoden:** Die Verwendung von natürlichen antimikrobiellen Mitteln erscheint für Lebensmittel sicher zu sein. Diese Art von Verbindungen erfährt potentielle Anwendung als natürliches Konservierungsmittel in der Nahrungsmittelindustrie. Dieser Beitrag stellt einige Beispiele der Verwendung von antimikrobiellen Mitteln, wie Chitosan, Nisin und Pektin dar.

**Ergebnissen und Fazit:** Die natürlichen antimikrobiellen, in den aktiven Verpackungen verwendeten Substanzen beseitigen zugunsten der Verbraucher die Gefahr der Migration von chemischen Stoffen zu Lebensmitteln.

**Codewörter:** Logistik, antibakterielle Verpackungen, natürliche antibakterielle Substanzen

---

Renata Dobrucka

Department of Industrial Products Quality and Ecology

Faculty of Commodity Science

Poznan University of Economics

al. Niepodległości 10, 61-875 Poznan, **Poland**

e-mail: [renata.dobrucka@ue.poznan.pl](mailto:renata.dobrucka@ue.poznan.pl)